

Copper 1,4,8,11-tetraazacyclotetradecane- N,N',N'',N'''-tetraacetic acid-octreotide Cu-TETA-OC

Created: December 20, 2004

Updated: March 14, 2005

Chemical name:	Copper 1,4,8,11-tetraazacyclotetradecane-N,N',N'',N'''-tetraacetic acid-octreotide	Structure available soon in PubChem [http://pubchem.ncbi.nlm.nih.gov/].
Abbreviated name:	Cu-TETA-OC; ^{64}Cu -TETA-OC; ^{64}Cu -TETA-octreotide; ^{64}Cu -TETA-OC	
Synonym:		
Backbone:	Peptide	
Target:	Somatostatin receptor	
Mechanism:	Binding of the octreotide	
Method of detection:	PET	
Source of signal:	^{64}Cu	
Activation:	No	
<i>In vitro</i> studies:	Yes	
Rodent studies:	Yes	
Other non-primate mammal studies:	Yes	
Non-human primate studies:	Yes	
Human studies:	Yes	

Background

[PubMed]

Somatostatin is a tetradecapeptide acting as an inhibitor of the release of somatotropin, glucagon, gastrointestinal hormones, and other secretory proteins. The targeting of somatostatin receptors with radiolabeled peptides has led to the development of a variety of agents for both diagnostic imaging and radiotherapy of somatostatin receptor-positive tumors, an area of cancer research where considerable progress has been made over the last few years.

^{64}Cu -TETA-octreotide (or ^{64}Cu -TETA-OC) is a somatostatin receptor showing high affinity for binding, both *in vitro* and *in vivo* (1). Its high rate of lesion detection, favorable dosimetry, and clearance properties make it a promising agent for positron emission tomography (PET) imaging of neuroendocrine tumors in patients (2). ^{64}Cu -TETA-OC displays a similar affinity as ^{111}In -DTPA-octreotide, a clinically approved imaging agent for somatostatin receptor-positive tumors.

Several animal studies also showed the therapeutic value of ^{64}Cu -TETA-OC as a tumor growth inhibitor (3). The mechanism of the tumor cell killing process is still unclear and currently under investigation (4). Preliminary subcellular distribution studies suggest a possible role played by the localization of ^{64}Cu to the tumor cell nuclei, a result from the dissociation of the metal from macro-cyclic chelators *in vivo* (5), followed by trafficking of the radiometal to the cell nuclei (4).

Synthesis

[PubMed]

TETA-OC can be prepared following a procedure by Anderson et al. (3). Briefly, the OC is protected with a tert-butoxycarbonyl (Boc) group by reaction with $(\text{Boc})_2\text{O}$ in Me_2SO , and TETA \cdot 4HCl \cdot 4H $_2\text{O}$ is neutralized with 4.5 equivalents of aqueous LiOH. The N-terminal amine of Boc-protected OC is conjugated to one of the carboxylic acid moieties on TETA with HBTU in Me_2SO , using di-isopropylethylamine and hydroxybenzotriazole as catalysts.

^{64}Cu -TETA-OC can be synthesized using the method described by Bass et al. (6). This procedure involves diluting $^{64}\text{CuCl}_2$ with 0.1 M NH_4OAc , at pH 5.5, then adding to TETA-OC and adjusting the final volume to 1.0-1.5 ml with buffer. After a 60-min incubation at room temperature, ^{64}Cu -TETA-OC is purified using a Sep-Pak cartridge (7). By following this procedure, the radiochemical purity of ^{64}Cu -TETA is >90%, and the radiochemical purity of ^{64}Cu -TETA-OC is >95% (by high-performance liquid chromatography, HPLC) (8).

In Vitro Studies: Testing in Cells and Tissues

[PubMed]

In vitro studies on somatostatin receptor-positive AR42J rat pancreatic tumors and focused on the subcellular distribution of ^{64}Cu -TETA-OC showed a localization of substantial quantities of ^{64}Cu to the cell nucleus and mitochondria (4). This process was shown to be a main contributing factor in the tumor cell killing process (4). It was also suggested that the instability of the moiety of ^{64}Cu -TETA under biological conditions was the cause of the translocation of ^{64}Cu to the nucleus.

Wang et al. (4) observed that the majority of ^{64}Cu -TETA-OC was internalized in AR42J cells by receptor-mediated endocytosis. This internalization process increased steadily over a 24-h time period, with low amounts of cell surface-associated activity, suggesting a rapid turnover and recycling of SSTR2 receptors.

Using a procedure modified from the one by Zinn et al. (9), Wang et al. (4) showed that the percentage of surface-bound, cell-associated activity ranged from almost 7% to >16%, with the amount of surface-bound activity increasing at 24 h, and that the majority of ^{64}Cu -TETA-OC was internalized.

Animal Studies

Rodents

[PubMed]

^{64}Cu -TETA-OC was shown to inhibit the growth of somatostatin receptor-positive tumors in rats at doses exhibiting minimal toxicity. In one study, Anderson et al. (3) performed experiments on rats bearing palpable CA20948 pancreatic tumors by injecting them with either a single 15-mCi dose, a fractionated amount of 15 mCi given in two to three doses over 2-8 days, or control agents of buffer OC. Results showed that ^{64}Cu -TETA-OC could greatly inhibit the growth of pancreatic tumors at doses causing minimal toxicity. The only toxicity observed in treated rats was a decrease in the white blood cell count, a significant drop for rats treated by single injection, and a slight decrease (with rebound) for those receiving a fractionated dose treatment. Injection of fractionated doses appeared to be more effective than a single dose treatment, showing a 25% reduction in tumor growth rate compared with single-dose injections (and a 75% reduction for the buffer control group). Estimated absorbed doses of ^{64}Cu -TETA-OC to the tumor were between 465 and 540 rads. At those doses, tumor inhibition—and even tumor regression (for large tumors)—was observed; however, all tumors eventually re-grew.

Using a model of tumor-bearing Lewis rats, the estimated human absorbed doses to normal organs showed the bladder wall (1.12 rad/mCi) and the lower large intestine (0.86 rad/mCi) to be the primary and secondary critical organs; the human effective dose equivalent was found to be 0.21 rad/mCi (3). ^{64}Cu -TETA-OC and ^{111}In -DTPA-OC showed similar biodistributions in tumor-bearing rat models (7).

Rat studies performed by de Jong et al. (10) showed that altering the OC structure slightly (by substitution of a tyrosine for phenylalanine, for example) resulted in a better uptake of the peptide in receptor-rich tissues such as adrenals, pancreas, pituitary, and tumor. However, rat studies showed retention of the activity of ^{64}Cu -TETA-OC in the blood, liver, and bone marrow, suggesting a possible dissociation of ^{64}Cu from TETA *in vivo*. In their study, Bass et al. (6) showed that ^{64}Cu dissociated from ^{64}Cu -TETA-OC and bound to proteins in large concentrations, such as superoxide dismutase (6).

Other Non-Primate Mammals

[PubMed]

No reference currently available.

Non-Human Primates

[PubMed]

PET imaging studies using ^{64}Cu -TETA-OC have been performed using non-human primates to estimate human absorbed doses. Data obtained by Anderson et al. (2) for ^{64}Cu -TETA-OC PET imaging on baboons showed the dose-limiting organs to be the bladder wall (0.62 rad/mCi), followed

by the kidneys (0.49 rad/mCi). The estimated human absorbed dose for the total body was found to be 0.07 rad/mCi. The large discrepancy between dosimetry in rats and baboons obtained for the intestinal absorbed doses was explained by the very different excretion patterns of these animals.

Human Studies

[PubMed]

Human absorbed doses of ^{64}Cu -TETA-OC to normal organs were estimated from biodistribution data in both tumor-bearing Lewis rats and baboons (2) (see sections on Rodents and Non-Human Primates).

Anderson et al. (2) performed ^{64}Cu -TETA-OC PET studies on eight patients with histologically proven neuroendocrine tumors (five with carcinoid tumors of the gastrointestinal tract and three with pancreatic islet cell tumors). Pharmacokinetic analysis of blood samples obtained from patients showed that ^{64}Cu -TETA-OC PET cleared rapidly from the blood. However, $7.9 \pm 3.7\%$ injected dose (ID) remained (range, 3.2–13.5 %ID) 4 h after injection. The activity decreased further from 6 to 22 h, with amounts ranging from 0.8 to 6.6 %ID (mean, $3.3 \pm 2.3\%$ ID). Large variations were observed from patient to patient. Similarly to the results obtained in rat studies, ^{64}Cu was retained in the blood, and ^{64}Cu -TETA-OC did not completely clear from the circulation (7).

Comparative studies between ^{64}Cu -TETA-OC PET and ^{111}In -DTPA-OC scintigraphy showed that, in general, more lesions were detected using ^{64}Cu -TETA-OC because of the higher resolution obtained with PET imaging. Nevertheless, the image quality was in some cases superior by using DTPA because of the absence of intense activity in the bladder and kidneys when using ^{111}In -DTPA-OC (2).

References

1. Lewis JS, Lewis MR, Srinivasan A, Schmidt MA, Wang J, Anderson CJ. Comparison of four ^{64}Cu -labeled somatostatin analogues in vitro and in a tumor-bearing rat model: evaluation of new derivatives for positron emission tomography imaging and targeted radiotherapy. *J Med Chem* 42:1341–1347; 1999. (PubMed)
2. Anderson CJ, Dehdashti F, Cutler PD, Schwarz SW, Laforest R, Bass LA, Lewis JS, McCarthy DW. ^{64}Cu -TETA-octreotide as a PET imaging agent for patients with neuroendocrine tumors. *J Nucl Med* 42:213–221; 2001. (PubMed)
3. Anderson CJ, Jones LA, Bass LA, Sherman EL, McCarthy DW, Cutler PD, Lanahan MV, Cristel ME, Lewis JS, Schwarz SW. Radiotherapy, toxicity and dosimetry of copper-64-TETA-octreotide in tumor-bearing rats. *J Nucl Med* 39:1944–1951; 1998. (PubMed)
4. Wang M, Caruano AL, Lewis MR, Meyer LA, van der Waal RP, Anderson CJ. Subcellular localization of radiolabeled somatostatin analogues: implications for targeted radiotherapy of cancer. *Cancer Res* 63:6864–6869; 2003. (PubMed)
5. Fjalling M, Andersson P, Forssell-Aronsson E, Gretarsdottir J, Johansson V, Tisell LE, Wangberg B, Nilsson O, Berg G, Michanek A, et al. Systemic radionuclide therapy using indium-111-DTPA-d-Phe1-octreotide in midgut carcinoid syndrome. *J Nucl Med* 37:1519–1521; 1996. (PubMed)
6. Bass LA, Wang M, Welch MJ, Anderson CJ. In vivo transchelation of copper-64 from TETA-octreotide to superoxide dismutase in rat liver. *Bioconjug Chem* 11:527–532; 2000. (PubMed)
7. Anderson CJ, Pajean TS, Edwards WB, Sherman EL, Rogers BE, Welch MJ. In vitro and in vivo evaluation of copper-64-octreotide conjugates. *J Nucl Med* 36:2315–2325; 1995. (PubMed)

8. Siegel BA, Dehdashti F, Mutch DG, Podoloff DA, Wendt R, Sutton GP, Burt RW, Ellis PR, Mathias CJ, Green MA, et al. Evaluation of ^{111}In -DTPA-folate as a receptor-targeted diagnostic agent for ovarian cancer: initial clinical results. *J Nucl Med* 44:700–707; 2003. (PubMed)
9. Zinn KR, Chaudhuri TR, Buchsbaum DJ, Mountz JM, Rogers BE. Simultaneous evaluation of dual gene transfer to adherent cells by gamma-ray imaging. *Nucl Med Biol* 28:135–144; 2001. (PubMed)
10. de Jong M, Bakker WH, Breeman WA, Bernard BF, Hofland LJ, Visser TJ, Srinivasan A, Schmidt M, Behe M, Macke HR, et al. Pre-clinical comparison of [DTPA0] octreotide, [DTPA0,Tyr3] octreotide and [DOTA0,Tyr3] octreotide as carriers for somatostatin receptor-targeted scintigraphy and radionuclide therapy. *Int J Cancer* 75:406–411; 1998. (PubMed)